

### Effect of *Solanum aculeastrum* on hematological parameters of Al-bino mice infected with *Aspergillus fumigatus*

Sara Ghalib Allwi Al-Saffy<sup>1</sup> and Dalia Abdalkareem Abdulshaheed<sup>2\*</sup>

<sup>1</sup>Department of Microbiology, College of Veterinary Medicine, University of Baghdad, Baghdad, Iraq; saragh199577@gmail.com. <https://orcid.org/0000-0002-6658-1237>

<sup>2</sup>Department of Microbiology, College of Veterinary Medicine, University of Baghdad, Baghdad, Iraq; dalia@covm.uobaghdad.edu.iq. <https://orcid.org/0000-0003-1369-8826>

\* Correspondence: dalia@covm.uobaghdad.edu.iq.

Available from: <http://dx.doi.org/10.21931/RB/2024.09.01.48>

#### ABSTRACT

The goal of the current study was to research the changes in hematological parameters: WBC count, RBCs count, Hb, PCV, neutrophil, lymphocyte, and monocyte in albino mice infected with *Aspergillus fumigatus* by intraperitoneal injection after induced immunosuppression by intraperitoneal injection of cortisone. The current research also examined an attempt to reduce the infection load by treating with *Solanum aculeastrum*. The result shows higher decreased significance ( $P \leq 0.05$ ) in RBCs, Hb, and PCV after being infected with *A. fumigatus*  $7.1 \pm 0.8$ ,  $11.3 \pm 0.5$  and  $41.5 \pm 2.4$ , respectively, while the total WBC count, neutrophil, lymphocyte, and monocytes were increased significantly ( $P \leq 0.05$ ) after treatment with *S. aculeastrum* in groups infected with *A. fumigatus*, compared to other groups. According to these results, we conclude that the alcoholic extract of *S. Astrum* has significant therapeutic and antifungal characteristics that lead to an increase in the total WBC count and, therefore, is considered a necessary alternative therapy for increasing immunity.

**Keywords:** Cortisone, Hematology, Fungi, Iraq.

#### INTRODUCTION

Fungi are a major, diverse, ubiquitous group of heterotrophic organisms that live as saprophytes or parasites or are related to many other organisms as symbiotes<sup>[1]</sup>. According to estimates of global richness, it constitutes the second largest group of organisms, with about 3 million species expected. Also, it ranks third among the eukaryotic kingdoms regarding the wealth of known species<sup>[2, 3]</sup>. The most prevalent pathogenic species in the animal kingdom is *Aspergillus fumigatus*, a saprotrophic fungus that lives vegetatively on decaying organic matter in the soil and spreads by asexual sporulation<sup>[4, 5]</sup>. This fungus can survive high temperatures above (50°C) and may happen in piles of decomposing plant matter. The fungus releases a significant amount of asexual airborne spores<sup>[6]</sup>. These fungi have a broad clinical spectrum, ranging from allergy to chronic infections and acute invasive aspergillosis (I.A.) in humans and animals<sup>[7]</sup>. Aspergilli are known for their capacity to secrete different types of biologically active chemical compounds, including antibiotics, mycotoxins, immune-suppressant substances, and cholesterol-lowering factors<sup>[8]</sup>. According to the host's underlying immunological state, these fungi can cause overlapping chronic, noninvasive types of infection that range from the formation of a fungal ball (Aspergilloma) to a long-lasting inflammatory and fibrotic process that is presently categorized as chronic lung infection similar to invasive pulmonary aspergillosis (IPA)<sup>[4, 9]</sup>.

*Solanum aculeastrum*, "goat bitter-apple" Hnzal (Arabic), is extensively dispersed in a native of Africa and South Africa. *S. aculeastrum* is a spiny perennial that grew to 3 m tall with white blooms and bears berries resembling lemons and turning yellow-green when ripe. For both people and domestic animals, the tart fruits of this plant are utilized medicinally in various techniques for the treatment of cancer, indigestion, and stomach disruption; the boiling extract of the fruits and leaves is administered orally route. Both fresh and cooked fruit are used as a therapy for acne, gonorrhea, and jigger wounds<sup>[10]</sup>, according to the presence of bioavailable

phyto-constituents including steroidal saponins, steroidal alkaloids, terpenes, flavonoids, lignans, sterols, phenolic compounds, and coumarins. *Solanum* spp. is essential in the nutraceutical and pharmaceutical industries. Both steroidal alkaloids and glycoalkaloids serve as important chemical indicators of this genus. Both ancient and modern systems of medicine place special importance on steroid alkaloids and glycoalkaloids since they exhibit a variety of bioactivities, including antibacterial, analgesic, immunomodulatory, hepatoprotective, neurogenetic, anticancer, etc. [11]. Considering the above facts and minimal studies, this study was designed to study the effects of *S. aculeastrum* on hematological parameters in albino mice infected with *A. fumigatus*.

---

## MATERIALS AND METHODS

### *Fungal isolation*

The fungus was isolated from different veterinary clinics and stray cats in Baghdad province, identified on Sabouraud Dextrose Agar and malt extract agar, and diagnosed by traditional morphological examination laboratory methods.

### *Preparation of A. fumigatus fungal suspension*

The suspension of *A. fumigatus* was prepared according to a previously carried out study [12].

### *Solanum aculeastrum fruit extraction*

*Solanum aculeastrum* (Hanzal) was purchased from local Baghdad markets, and the extraction was performed previously [13].

### *Induction of immunosuppression*

Immunosuppression was induced by treatment of the mice with cortisone intraperitoneal administration at a single dose one day before conidial administration. Each mouse receives 2mg of Dexamethasone per mouse (14).

### *Animal and design experiments*

Fifty-six albino mice were used in the study at 10-12 weeks and weighing  $25.3 \pm 0.9$  g, maintained on a standard laboratory diet, water and temperature-controlled at the animal house laboratory at the College of Veterinary Medicine (University of Baghdad, Baghdad, Iraq), were randomly assigned to different groups: 8 mice per group as following:

- G1: Negative control mice (untreated).
- G2: Infected mice with a single dose of *A. fumigatus* at  $2 \times 10^7$  cells/ml per mouse intraperitoneally (I/P).
- G3: Treated mice with cortisone at 2 mg/mice I/P.
- G4: Treated mice orally with a single dose of *S. aculeastrum* at 10 mg/kg B. W.
- G5: In which mice were treated with a single dose of cortisone at 2mg/mice I/P for one day prior to fungal spore infection and then infected with a single dose of *A. fumigates*  $2 \times 10^7$  cells/ml per mouse I/P.
- G6: In which mice were treated with a single dose of *S. aculestrum* at 10 mg/kg B. W orally, one week after infection with a single dose of *A. fumigates*  $2 \times 10^7$  cells/ml per mouse I/P.
- G7: Mice were treated orally with a single dose of *S. aculestrum* at 10 mg/kg B. W simultaneously with the fungal infection.

### *Blood samples collection*

According to (16), the blood samples from all groups were taken at the end of the experiment.

### *Statistical analysis*

SPSS calculated data for Windows TM version 23. 0 by using one-way ANOVA. All experimental data are presented as Mean  $\pm$  S.E. and significant differences at  $P \leq 0.05$  [17].

## RESULTS

### Hematological study

Hematological parameters: total WBC counts, neutrophils, lymphocytes, monocytes, RBC count, Hb level, and PCV. The blood samples were taken from all the groups to analyze by employment tubes containing an anticoagulant agent in the laboratory. In the current study, the results showed that the control positive group of *A. fumigatus* infection, cortisone and group infected with *A. fumigatus* plus cortisone treated caused significant ( $p \leq 0.05$ ) decrease in RBCs count, Hb and PCV, whereas, no significant ( $p \leq 0.05$ ) differences were noticed in groups of mice treated with *Solanum aculeastrum* alone or plus cortisone and *A. fumigatus* in both G6 and G7 when compare with G1 (Table 1).

Parameter	G1	G2	G3	G4	G5	G6	G7
RBCs $10^6/\text{mm}^3$	9.9 ± 0.4 a	7.1 ± 0.8 b	7.8 ± 0.3 b	9.7 ± 1.3 a	7.9 ± 0.1 b	9.9 ± 0.3 a	9.7 ± 0.5 a
Hb mg/dl	14.2 ± 0.1 1a	11.3 ± 0.5 b	11.4 ± 0.3 b	14.4 ± 1.5 a	12.4 ± 0.6 b	13.7 ± 0.9 a	13.4 ± 0.3 a
PCV %	46.8 ± 0.2 2a	41.5 ± 2.4 b	39.9 ± 0.4 b	44.7 ± 6.3 a	43.6 ± 1.5 b	43.5 ± 1.8 a	47.4 ± 2.4 a

Variation in horizontal small letters refers to significant differences ( $P < 0.05$ ).

**Table 1: Effects of *A. fumigatus* infection, cortisone, and *S. aculeastrum* treated after two weeks on parameters of RBC count, Hb, and PCV (mean ± S.E.).**

Total WBCs, neutrophils, lymphocytes, and monocyte count were found to be decreased significantly ( $p \leq 0.05$ ) in both groups infected with *A. fumigates* and *S. aculeastrum* alone. Also, mice representing control-positive cortisone decreased significantly ( $p \leq 0.05$ ) in neutrophils and monocytes. At the same time, total WBCs and lymphocytes were not affected significantly ( $p \leq 0.05$ ). The results also showed no significant differences ( $p \leq 0.05$ ) in a group of mice treated with cortisone plus infection of *A. fumigatus*. Total WBCs and lymphocytes were increased significantly ( $p \leq 0.05$ ), and neutrophils and monocytes were not affected significantly ( $p \leq 0.05$ ) in G6 treated with *S. aculeastrum* after one week of infection. Total WBCs, neutrophils, and lymphocytes were increased significantly ( $p \leq 0.05$ ), and monocytes were not affected significantly ( $p \leq 0.05$ ) in the group treated with *S. aculeastrum* at the same time of infection comparable with the controls regarding the WBCs (Table 2).

Parameter	G1	G2	G3	G4	G5	G6	G7
WBCs $10^9/\text{L}$	5.3 ± 0.5 a	3.7 ± 0.8 b	5.0 ± 0.5 a	2.5 ± 0.4 b	5.8 ± 1.9 a	6.6 ± 0.9 a	7.6 ± 3.0 b
Neutrophil %	3.2 ± 0.8 a	1.8 ± 0.7 b	1.8 ± 0.1 b	1.6 ± 0.5 b	2.4 ± 0.9 a	3.4 ± 1.1 a	4.2 ± 1.0 b
Lymphocyte %	2.1 ± 0.4 a	1.4 ± 0.2 b	2.6 ± 0.4 a	1.1 ± 0.5b	2.5 ± 0.8 a	1.5 ± 0.2 b	4.0 ± 0.3 b
Monocyte %	1.2 ± 0.1 a	0.1 ± 0 b	0.1 ± 0 b	0.1 ± 0.5 b	0.2 ± 0 a	0.6 ± 0.1 a	0.7 ± 0.04 a

Variation in horizontal small letters refers to significant differences ( $P < 0.05$ ).

**Table 2: Effects of *Aspergillus fumigatus* infection, cortisone, and *Solanum aculeastrum* treated after two weeks on parameters of (WBCs, neutrophils, lymphocytes, and monocytes) (mean ± S.E.).**

## DISCUSSION

The present results indicated that the hematological picture of *A. fumigatus*-infected mice is similar to that of another study [18]. The mycotoxin from pathogenic fungi caused anemia, observed as decreases in PCV Hb% and RBC count. Also, the phospholipase enzyme in *Aspergillus* spp. is the essential virulence factor that can cause destabilization and penetration, break down membrane phospholipids surrounding the red blood cell, and generate arachidonic acid [19]. Additionally, phospholipase hydrolyzes RBCs to release phosphatidylserine and produce lysophosphatidic acid (LPA), the latter of which results in the flow of substances through a blood cell membrane and causes swelling before exploding [20].

Previous studies also found decreased hematological parameters because of cortisone treatment [21, 22]. Exposure to Dexamethasone demonstrates a significant change in red blood cells, such as anemia, destruction of cells, or inhibition of hematopoiesis. All these reasons affect RBC, Hb, and PCV levels, and these changes led to observing the result compared with a control group and explaining the result that Dexamethasone may cause suppression of the bone marrow. Such decreases could also be attributed to hemodilution. However, they could also result from RBC reaction or inhibition of RBC synthesis, which limits the capacity of these cells to absorb oxygen in conjunction with increased hemolysis brought on by excessive physical stress and results in severe anemia. The WBC decrease results in the lysis of neutrophils affected by phospholipase enzymes, the elimination of lipid phosphorus, and hydrolysis of membrane phospholipids occurred after exposure to this enzyme. This led to decreased cell size, sphering, and increased susceptibility to osmotic stress, which altered the cell's functional properties and caused cell lysis [23]. Another reason represented that phospholipase enzyme and *Aspergillus* spp. The infection effect on hormones responsible for the production of blood cells is represented by the erythropoietin hormone from the kidney, which is the primary catalyst for the production of blood cells; this finding agrees with other researchers [24].

The reduction in neutrophil and monocyte counts after cortisone-treated mice indicated immunosuppression. This finding agrees with another study [25]. Steroids impair alveolar macrophage activity, lowering the primary defense against lung infection. Additionally, they affect T and B cell lymphocytes and reduce cytokine production, which impairs the adaptive immune response to invasive aspergillosis [26].

*Solanum aculeastrum* affects hematological parameters; the results indicated that plants cause a decrease in total WBC count and differential cells. WBC count is an indicator of an organism's capacity to eradicate infection. A reduction in WBC count in the group of mice treated with *Solanum aculeastrum* agrees with another study [27]. The animal's capacity to fight infection, attack, and destroy infectious agents in the blood may be adversely impacted by the decline in WBC levels. Additionally, the immune system's effector cells may have a severe effect. Also, the WBC count decreases, as observed by other results [28]. The decrease in WBC count suggests that the extract ingredients may have damaged or prevented the maturation of these blood cells. The committed stem cells that produce these blood cells are regulated by proliferation, differentiation, and maturation by granulocyte-macrophage colony-stimulating factors, interleukins IL-2, IL-4, and IL-5. Therefore, the extract might have interfered with the sensitivity of the committed stem cells responsible for generating these white blood cells and differential cells, or it might have decreased the synthesis of these regulatory factors [29-32].

Mice in both G6 and G7 showed an increase in total WBC count and its differential cells compared with other treated groups; best results were obtained when mice treated at the same time of infection compared with mice received the plant extract after one week of fungal spores infected, the stress factor of infection on the immune system and weakness of mice body can interfere with immune responses. Also, lymphocyte counts may be decreased with handling or other stressors and with age. Increased neutrophil counts (neutrophilia) are commonly seen in conditions of infection and acute inflammation and are related to immune system reactions to stress or excitement and infectious diseases [33].

Previous research proved that *S. aculeastrum* has a chemical and pharmaceutical component that acts as an antifungal against a number of fungal species [34, 35]. Other researchers have investigated the effect of ethanolic extract using the diffusion method, and the findings showed that the strongest antifungal activity was by *A. fumigatus* [13, 36, 37]. At the same time, other authors found that alcohol extracts from fruit ultimately impeded the growth of *A. flavus* (100%) [28, 35-37]. The results of the present study agree with all the research above; the hematological findings showed improvements in white blood cells, neutrophils, lymphocytes and monocytes when given the ethanolic extract orally at the same time as infected with *A. fumigatus*, indicating the treatment power of the plant at the applied dose of 10 mg/kg B. W was elevated the immune status by increasing of immune cells that fight infection comparing with other groups.

---

## CONCLUSIONS

This study concluded that using the alcoholic extract of *S. aculeastrum* could provide an effective therapeutic tool in the treatment of fungal infections as well as in the correction of abnormalities in hematological and immunological markers due to exposure to different infections. Furthermore, studies are of great importance to detect the efficacy of this extract on body organs and other infections.

**Author Contributions:** Conceptualization, S. G. A. A. and D. A. A.; methodology, S. G. A. A. and D. A. A.; software, S. G. A. A.; validation, S. G. A. A. and D. A. A.; formal analysis, S. G. A. A. and D. A. A.; investigation, S. G. A. A.; resources, S. G. A. A.; data curation, D. A. A.; writing-original draft preparation, S. G. A. A.; writing-eview and editing, S. G. A. A. and D. A. A.; visualization, D. A. A.; supervision, D. A. A.; project administration, S. G. A. A. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research received no external funding.

**Institutional Review Board Statement:** The study was conducted according to the guidelines of the Declaration of Helsinki and approved by the Scientific Ethics Committee of the Department of Microbiology at the College of Veterinary Medicine (University of Baghdad, Baghdad, Iraq).

**Acknowledgments:** The authors dramatically acknowledge the Head Department and the staff of the Department of Microbiology at the College of Veterinary Medicine for their valuable support.

**Conflicts of Interest:** The authors declare no conflict of interest.

---

## REFERENCES

1. McCoy, C.W; Samson, R.A; Boucias, D.G. Entomogenous fungi. Handbook of natural pesticides, *CRC Press*, **2019**, pp.151-236.
2. Barkai-Golan, R.; Paster, N. Mycotoxins in fruits and vegetables. *Elsevier*, **2011**, pp. 74.
3. Lücking, R.; Aime, M.C.; Robbertse, B.; Miller, A.N.; Ariyawansa, H.A.; Aoki, T.; Latgé, J.P.; Chamilos, G. *Aspergillus fumigatus* and Aspergillosis in 2019. *Clin Microbiol Rev* **2019**, 33(1), 140-148.
4. Tell, L.A.; Burco, J.D.; Woods, L.; Clemons, K.V. Aspergillosis in birds and mammals: considerations for veterinary medicine. In *Recent Developments in Fungal Diseases of Laboratory Animals*. Springer, Cham. **2019**. pp.49-72.
5. Schoustra, S.E.; Debets, A.J.; Rijs, A.J.; Zhang, J.; Snelders, E.; Leendertse, P.C.; Verweij, P.E. Environmental hotspots for azole resistance selection of *Aspergillus fumigatus*, the Netherlands. *Emerg Infect Dis* **2019**, 25 (7), 1347-1353.
6. Resendiz Sharpe, A.; Lagrou, K.; Meis, J.F.; Chowdhary, A.; Lockhart, S.R.; Verweij, P.E.; ISHAM/ECMM *Aspergillus* Resistance Surveillance Working Group. Triazole resistance surveillance in *Aspergillus fumigatus*. *Med Mycol* **2018**, 56(suppl\_1), S83-S92.
7. May, G.S. Mitogen-activated protein kinase pathways in *Aspergilli*. In *The aspergilli*. *CRC Press*. **2007**, pp.141-148.
8. Dewi, I.M.; Janssen, N.A.; Rosati, D.; Bruno, M.; Netea, M.G.; Brüggemann, R.J.; van de Veerdonk, F.L. Invasive pulmonary aspergillosis associated with viral pneumonitis. *Curr Opin Microbiol* **2021**, 62, 21-27.
9. Koduru, S.; Grierson, D.S.; Afolayan, A.J. Antimicrobial Activity of *Solanum aculeastrum*. *Pharm Biol* **2006**, 44(4), 283-286.
10. Patel, P.; Prasad, A.; Srivastava, K.; Singh, S.S.; Chakrabarty, D.; Misra, P. Updates on steroidal alkaloids and glycoalkaloids in *Solanum* spp.: Biosynthesis, in vitro production and pharmacological values. *Stud Nat Prod Chem* **2021**, 69, 99-127.
11. Su, H.; Li, C.; Wang, Y.; Li, Y.; Dong, L.; Li, L.; Zhu, M. Kinetic host defense of the mice infected with *Aspergillus Fumigatus*. *Future Microbiol* **2019**, 14(8), 705-716.

12. Eidi, S.; Azadi, H.G.; Rahbar, N.; Mehmannaavaz, H.R. Evaluation of antifungal activity of hydroalcoholic extracts of *Citrulluscolocynthis* fruit. *J Herb Med* **2015**, *5* (1), 36-40.
13. Marina A.Shevchenko, Andrey O.Bogorodskiy, Natalia I.Troyanova, Ekaterina A.Servuli, Elena L.Bolkhovitina, Georg Büldt, ChristophFahlke, Valentin I. Gordeliy, Thomas Gensch, Valentin I. Borshchevskiy, Alexander M. Sapozhnikov, "Aspergillus fumigatus Infection-Induced Neutrophil Recruitment and Location in the Conducting Airway of Immunocompetent, Neutropenic, and Immunosuppressed Mice. *J Immunol Res* **2018**, *12*, 20-28.
14. Gargiulo, S.; Greco, A.; Gramanzini, M.; Esposito, S.; Affuso, A.; Brunetti, A.; Vesce, G. Mice anesthesia, analgesia, and care, Part I: anesthetic considerations in preclinical research. *ILAR J* **2012**, *53*(1), E55-E69.
15. Coles, E.H. *Veterinary clinical Pathology* 4th ed WB Saunders company Philadelphia.London, Toronto, Mexico, Riodejenario, Sydney, Tokyo Hong Kong, **1986**, 136-170.
16. Gharban, H.A. Cumulative Effect of Subclinical Mastitis on Immunological and Biochemical Parameters in Cow Milk.*Arch Razi Instit*, **2021**, *76* (6), 1599-1608.
17. Mohamad, S.H.; Thalij, K.M.; AL-Bander, K.; Dheeb, B.I. Effects of Allergic fungi on hematological and immunological parameters of human patients and rabbits. *Egypt Acad J Biol Sci GMicrobiol* **2014**, *6*(2), 41-48.
18. Mansoor, S. S.; Al-Esawi, J. S. . . ; Al-Falahi, M. N. Assessing The Efficiency Of Cement Kiln Dust For Heavy Metals Removal From Simulated Polluted Water. *JLSAR* **2023**, *4*, 45-52.
19. Neidlinger, N.A.; Larkin, S.K.; Bhagat, A.; Victorino, G.P.; Kuypers, F.A. Hydrolysis of phosphatidylserine-exposing red blood cells by secretory phospholipase A2 generates lysophosphatidic acid and results in vascular dysfunction. *J Biol Chem* **2006**, *281* (2), 775-781.
20. Razzaq, S.A.; Jaber, I.J.; Kadhim, S.A.; Abbas, A.A. Pharmacological Effects of Dexamethasone in Rats. *Indian J Forensic Med Toxicol* **2020**,*14* (3), 1003.
21. Ribas, J.L.C.; Zampronio, A.R.; Silva De Assis, H.C. Effects of trophic exposure to diclofenac and Dexamethasone on hematological parameters and immune response in freshwater fish. *Environ Toxicol Chem* **2016**, *35*(4), 975-982.
22. Ghannoum, M.A. Potential role of phospholipases in virulence and fungal pathogenesis. *Clin Microbiol Rev* **2000**,*13* (1), 122-143.
23. Auda, M.A. Effect of phospholipase and the fungus which it produced (*Aspergillusniger*) on the some of blood parameters of male mice (*Musmusculus*). *Al-Qadisiyah J Pure Sci* **2009**, *14* (1). 22-39.
24. Al-Maliki, S.J.; Al-Ali, A.A.; Kathim, A.S. Effect of corticosteroids cortisol hormone [hydrocortisone] on the of the blood parameter in pregnant and non-pregnant laboratory females mice. *J Histol Cell Biol* **2018**, *1* (1): 16-22.
25. S. Hameed, T., Sawicka, B. Role Of Agricultural Extension In Adoption Of Sustainable Agriculture Practices. *Anbar Journal Of Agricultural Sciences*, **2023**; *21*(1): 250-260. doi: 10.32649/ajas.2023.179947.
26. A A Al-Azzami , Th T Mohammed . Effect of Adding Dry Leaves of Lemongrass (*Cymbopogon Citratus*) To the Diet on Some Biochemical Tests of Blood in Broiler (Ross 308). IOP Conf Ser Earth Environ Sci 2023, 1252 (1), 12125. <https://doi.org/10.1088/1755-1315/1252/1/012125>.
27. Aboyade, O.M.; Yakubu, M.T.; Grierson, D.S.; Afolayan, A.J. Studies on the toxicological effect of the aqueous extract of the fresh, dried and boiled berries of *SolanumaculeastrumDunal* in male Wistar rats.*Hum Exp Toxicol* **2009**, *28* (12), 765-775.
28. Aboyade, O.M.; Yakubu, M.T.; Grierson, D.S.; Afolayan, A.J. Safety evaluation of aqueous extract of unripe berries of *Solanumaculeastrum* in male wistar rats. *Afr J Pharm Pharmacol* **2010**, *4*(3), 90-97.
29. Al-gharban, H.A.; Dhahir, S.H. Serological diagnosis of persistent infection with *Anaplasma marginale* bacteria in cattle. *Iraqi J Vet Med* **2015**, *39* (1), 33-39
30. Gharban, H.A.; Yousif, A.A. Serological and Molecular Phylogenetic Detection of *Coxiella burnetii* in Lactating Cows, Iraq. *Iraqi J Vet Med* **2020**, *44*(E0), 42-50
31. Gharban, H.A.; Yousif, A.A. Serological, Clinical and Hematological prevalence of *Coxiella burnetii* in Adult Cows, Iraq. *Biochem Cell Arch* **2020**, *20*(1), 67-74.
32. Ameen M. Shaman , Th. T. Mohammed. Effect Using Feed Additives Instead of Imported Premixes Affects the Physiology of Broiler Chickens. IOP Conf Ser Earth Environ Sci 2023, 1262 (7), 72080. <https://doi.org/10.1088/1755-1315/1262/7/072080>.

33. A A Al-Azzami , Th T Mohammed . The Effect of Adding Lemongrass Leaf Powder (*Cymbopogon Citratus*) to the Diet as a Natural Supplement on Some Productive Traits and Oxidation Indicators in Broiler (Ross 308). IOP Conf Ser Earth Environ Sci 2023, 1252 (1), 12123. <https://doi.org/10.1088/1755-1315/1252/1/012123>.
34. O'Connell, K.E.; Mikkola, A.M.; Stepanek, A.M.; Vernet, A.; Hall, C.D.; Sun, C.C.; Brown, D.E. Practical murine hematopathology: a comparative review and implications for research. *Comp Med* **2015**, 65(2), 96-113.
35. Gurudeban, S.; Ramanathan, T.; Satyavani, K.; Dhinesh, T. Antimicrobial effect of coastal medicinal plant "Citrulluscolocynthis against pathogenic microorganisms. *Afr J Pure Appl Chem* **2011**, 5(5), 119-122.
36. Hameed, B.; Ali, Q.; Hafeez, M.M.; Malik, A. Antibacterial and antifungal activity of fruit, seed and root extracts of *Citrulluscolocynthis* plant. *Biol Clin Sci Res J* **2020**, 19 (1), 1-15.
37. Mohammad, H.A.; Ajaj, E.A.; Gharban, H.A. The first study on confirmation and risk factors of acute and chronic canine distemper in stray dogs in Wasit Province, Iraq, using enzyme-linked immunosorbent assay and reverse transcription-polymerase chain reaction. *Vet World* **2022**, 15 (4), 968-974
38. Koduru, S.; Grierson, D.S.; Afolayan, A.J. Antimicrobial Activity of *Solanumaculeastrum*. *Pharm Biol* **2006**, 44(4), 283-286.

**Received:** October 9th 2023/ **Accepted:** January 15th 2024 / **Published:** 15 February 2024

**Citation:** Chango, M.; Rosero, G.; Erazo, N.; Álvarez, P. Effect of *Solanum aculeastrum* on hematological parameters of Albino mice infected with *Aspergillus fumigatus*. *Revis Bionatura* 2024; 9 (1) 48. <http://dx.doi.org/10.21931/RB/2024.09.01.48>

**Additional information** Correspondence should be addressed to [dalia@covm](mailto:dalia@covm).

**Peer review information.** Bionatura thanks anonymous reviewer(s) for their contribution to the peer review of this work using <https://reviewerlocator.webofscience.com/>

All articles published by Bionatura Journal are made freely and permanently accessible online immediately upon publication, without subscription charges or registration barriers.

**Bionatura ISSN.** First 13909355 Ecuador. **Scopus coverage years:** from 2016 to the present

**Publisher's Note:** Bionatura stays neutral concerning jurisdictional claims in published maps and institutional affiliations.

**Copyright:** © 2023 by the authors. They were submitted for possible open-access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).